

Chapter 1 1
Ordination Methods and the Evaluation 2
of Ediacaran Communities 3

Matthew E. Clapham 4

Contents

1.1 Introduction..... 000
 1.2 Dataset Summary..... 000
 1.3 Data Standardization..... 000
 1.4 Ordination Methods..... 000
 1.4.1 Principal Components Analysis (PCA)..... 000
 1.4.2 Correspondence and Detrended Correspondence
 Analysis (CA/DCA) 000
 1.4.3 Non-Metric Multidimensional Scaling (NMDS)..... 000
 1.5 Comparison and Interpretation of Results..... 000
 1.6 Conclusion..... 000
 References..... 000

Abstract Analysis of paleocommunity data poses a challenge because of its 5
 multivariate nature, containing counts of many species in many samples. Comparison 6
 of the abundance of a single species among all samples provides only incom- 7
 plete information, whereas attempting to consider every species is impractical. 8
 Ordination methods are analytical techniques that reduce the original multivariate 9
 dataset to a few important components by creating new synthetic variables designed 10
 to explain the maximum amount of original data variability. The ultimate goal is to 11
 order the samples along ecologically or environmentally meaningful gradients 12
 in order to interpret differences in community structure. This chapter describes 13
 three of the most widely-used ordination methods, principal components analysis 14
 (PCA), detrended correspondence analysis (DCA), and non-metric multidimensional 15
 scaling (NMDS), explaining the methodology of each and outlining their strengths 16
 and weaknesses for analysis of paleoecological data. The techniques are illustrated 17
 using Ediacaran paleocommunity data from Mistaken Point, Newfoundland. PCA 18
 relies on assumptions that are inappropriate for ecological data, such as the requirement 19

M.E. Clapham (✉)
 Department of Earth and Planetary Sciences, University of California,
 Santa Cruz, CA 95060, USA
 e-mail: mclapham@ucsc.edu

20 that species abundances change in a linear fashion along the environmental
21 gradient, and is not well suited for community ordination. In contrast, DCA and
22 NMDS both perform well with ecological data; DCA incorporates a more ecolog-
23 ically-realistic measure of distance between samples but some of the detrending
24 methods have been criticized, whereas NMDS only assumes a monotonic relation-
25 ship between compositional similarity and gradient distance. The two methods also
26 have complementary strengths, with DCA typically better at extracting the primary
27 gradient and NMDS better at resolving the overall pattern.

28 **Keywords**

[AU1]

29 **1.1 Introduction**

30 Ecologists and paleoecologists are commonly faced with large datasets containing
31 many variables from many samples (multivariate data), with the ultimate goal of
32 revealing and interpreting ecologically significant patterns in those data. In paleoecol-
33 ogy these datasets often take the form of an occurrence matrix of presence/absence
34 or relative-abundance counts of perhaps 10 to >100 taxa (variables) from as many as
35 10–100s of collection sites. Analysis of between-sample similarities in a single
36 variable (e.g., the abundance of a single taxon) is inadequate at capturing the full
37 range of variability in the original dataset. However, the original dataset also contains
38 a great deal of redundant information as, for example, several taxa can respond in a
39 similar fashion to the same environmental gradient (Palmer 1993). Thus, it is almost
40 always desirable to reduce the number of variables, typically to the two or three most
41 important components, allowing the data to be displayed and interpreted more easily
42 than by examining the dozens of variables in the original data.

43 A wide range of different analytical methods have been devised for reducing
44 complex multivariate datasets so that they can be displayed in two or three dimensions
45 (James and McCulloch 1990; Shi 1993). They are collectively called ordination tech-
46 niques and in ecology and paleoecology they typically are used for gradient analysis –
47 identifying environmental gradients and placing species in their correct position along
48 those gradients (Bray and Curtis 1957). Environmental gradients, such as depth (more
49 precisely, depth-related variables such as energy and substrate) in marine communities,
50 exert a fundamental control on the spatial distribution of species and, as a result, are the
51 primary influence on community composition (Cisne and Rabe 1978; Holland
52 et al. 2001; Holland 2005). Other important gradients may include biogeographic varia-
53 tion, other environmental changes such as salinity (in estuarine environments, for
54 example), temporal changes in community structure due to origination/immigration and
55 extinction/emigration, or ecological succession as argued by Clapham et al. (2003).

56 This chapter will focus on three techniques for reconstructing environmental
57 gradients and analyzing community structure: (1) Principal Components Analy-
58 sis (PCA), (2) Correspondence Analysis (CA) and its relative Detrended
59 Correspondence Analysis (DCA), and (3) Non-Metric Multidimensional

Scaling (NMDS). Other methods (Polar Ordination and Principal Coordinates Analysis) will be discussed briefly but are less commonly applied by paleo-ecologists. These are all indirect gradient analysis methods, as they use the distribution of species among the sites to infer the original environmental gradients that structured those sites (Minchin 1987; Palmer 1993). In contrast, direct gradient analysis techniques (e.g., Canonical Correspondence Analysis) explicitly relate species composition to measured environmental parameters (Palmer 1993), but are less applicable to paleontological data where environmental parameters typically are not directly measured or even estimated. The key differences between these three indirect gradient analysis methods are the mechanisms by which they quantify sample dissimilarity and the assumptions inherent in relating dissimilarity to separation along the environmental gradient (Faith et al. 1987). The implications of those differences will be discussed in greater detail below, with the ultimate goal of answering the question: which ordination method should you use to analyze your data?

1.2 Dataset Summary

This chapter will use a dataset of relative abundances from Ediacaran communities at Mistaken Point (Table 1.1), previously used by Clapham et al. (2003), to explore the procedures, assumptions, and application of different multivariate techniques (see Chap. 3 for a more detailed description of the fossil sites). The occurrence matrix contains abundance counts of 16 taxa from nine fossiliferous bedding planes, including three outcrops of the well-known “E” surface. All abundances

Table 1.1 Occurrence matrix showing abundance of 16 taxa in nine Mistaken Point samples

Sample/Taxon	PC	BC	LMP	D	Ey	Eq	Ewc	G	SH
<i>Aspidella</i>	0	0	0	1	3	0	1	0	1
<i>Bradgatia</i>	0	1	0	76	226	15	14	55	10
<i>Charnia</i>	18	0	152	20	84	5	14	4	1
“Charnia” B	0	26	58	0	0	0	0	0	0
<i>Charniodiscus</i>	0	0	13	0	1,398	52	35	49	3
“Dusters”	0	0	4	8	612	28	35	5	0
<i>Fractofusus</i> “Spindle”	0	76	4	1,169	1,497	71	100	0	0
<i>Hiemalora</i>	0	0	1	0	21	1	0	0	0
“Holdfast Frond”	0	0	0	0	0	0	0	22	0
<i>Ivesheadia</i> (“ <i>Ivesia</i> ”)	79	1	1	7	77	5	3	8	27
“Lobate Disc”	0	0	0	0	142	3	10	0	0
<i>Hapsidophyllas</i> “Network”	0	0	0	1	1	0	2	0	0
“Ostrich Feather”	0	0	62	0	0	0	0	0	0
<i>Pectinifrons</i> “Pectinate”	0	0	0	175	0	0	0	0	304
“Spoon Frond”	0	0	0	0	1	1	1	0	0
<i>Thectardis</i> “Triangle”	140	0	0	0	25	3	7	0	0
Total	237	104	295	1,457	4,087	184	222	143	346

82 were standardized to percent abundance by normalizing by the total number of
83 specimens per sample, as many of the analyses are sensitive to variations in abso-
84 lute abundance among sites that reflect sampling intensity rather than an actual
85 ecological parameter. Most taxa are identified to genus level but several are poly-
86 phyletic form taxa: the “dusters” category likely encompasses two or more undescribed
87 genera, while *Charnia* counts also include the recently-described genus *Beothukis*,
88 which was not differentiated in the original data collection of Clapham et al. (2003).
89 All of the ordination techniques will be performed on the same occurrence matrix to
90 facilitate understanding of their methodology and enable comparison of their results.
91 Most analyses shown here were performed with the free software package PAST
92 (Hammer et al. 2001) but can be done with a variety of statistical programs.

93 1.3 Data Standardization

94 The first important choice to be made, even before deciding on the appropriate
95 ordination technique, is whether to analyze the raw abundance counts or whether to
96 standardize them. All of the analyses in this chapter will be performed after converting
97 each raw abundance value to percent abundance, normalizing the data to eliminate
98 the influence of variations in total sample size. Otherwise, it is possible that two
99 samples may be erroneously grouped together because rare species in the large
100 sample have a similar raw abundance to common species in the small collection.
101 This standardization was chosen because sample size in the Mistaken Point dataset
102 is a reflection of sampling intensity, rather than having ecological significance. This is
103 the case in many, but not all, paleoecological datasets, and investigators will need to
104 consider whether variations in total abundance have biological significance (in which
105 case they should not convert the data to proportions) or reflect sampling intensity.

106 It is also possible to normalize species to their maximum abundance so that the
107 species relative abundances all sum to a value of 1 (or 100%), or to perform a double
108 standardization by both site and species totals (Faith et al. 1987). Standardization by
109 species total abundance strongly weights rare species and reduces the influence of abun-
110 dant taxa, which may not be appropriate unless the species occur at similar abundance
111 or different trophic levels are present and the investigator wishes to account for the rarity
112 of carnivores relative to primary producers, for example Jongman et al. (1995).

113 1.4 Ordination Methods

114 1.4.1 Principal Components Analysis (PCA)

115 Principal Components Analysis is one of the oldest techniques used in multivariate
116 data ordination and was the first to be applied to ecological data (Goodall 1954).

It still enjoys some use in paleoecology (e.g., Rodriguez 2004; Botquelen et al. 2006; McKinney and Hageman 2006) despite having several attributes that are not ideally suited to certain types of ecological data (Gauch and Whittaker 1972; Fasham 1977; Minchin 1987). The most severe issue, discussed in more detail below, is the distortion induced by fitting a linear (Euclidean) distance model to non-linear ecological gradients (Gauch and Whittaker 1972; Minchin 1987). As a result, PCA should not be used for analysis of ecological (species count) data. It is instructive, however, to examine the analytical procedure used in PCA because it illustrates the general principles of ordination, in particular eigenanalysis-based multivariate ordination. PCA, like all ordination, seeks to summarize the original multivariate dataset, which can often contain dozens of species at multiple sites, in a smaller number of new, synthetic variables – the “principal components” in the method’s name (James and McCulloch 1990). It is based on linear algebra, using a technique called eigenanalysis to rigidly rotate the original data matrix (i.e., changing the coordinate system without altering the relative position of the points) so that the new principal component axes account for the maximum amount of variability in the original data (Quinn and Keough 2002). The underlying mathematical basis is complex but the basics of PCA and other eigenanalysis methods can be explained graphically by considering a simplified example with only two variables (Fig. 1.1).

In this simple example, analysis of ecological data begins with a bivariate plot in which the axes are the abundance of species 1 and species 2, and the data points represent each sample in the study (Fig. 1.1a). The data are then “centered” by subtracting the mean species abundance from the abundance in each sample; this has the effect of moving the points so that the graph axes run through the mean of each species (Fig. 1.1b). The final step rotates the axes so that one axis, principal component 1, is aligned with the maximum variance of the sampled points (Fig. 1.1c). In fact, the new axes are simply obtained by a linear regression through the data points. Principal component 2 is created in a similar fashion and accounts for the maximum amount of remaining variance with the constraint that it is perpendicular to axis 1. This procedure can easily be conceptually generalized to

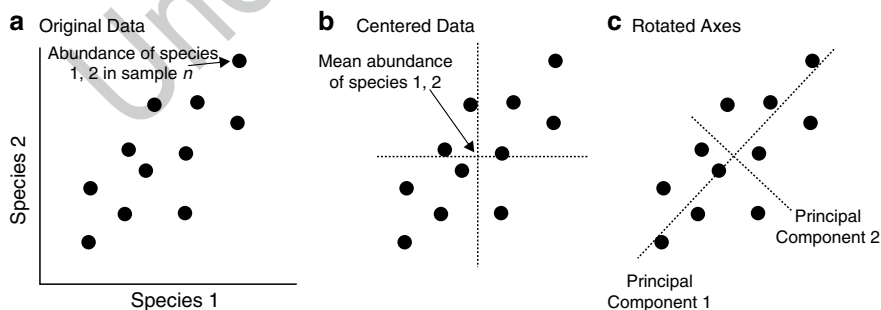


Fig. 1.1 Simplified two-variable example conceptually illustrating the process involved in PCA. (a) The abundance of each species is plotted on a bivariate (or multivariate) plot. (b) Data points are centered so that the axes pass through the mean point of each species. (c) Axes are rotated to align with the direction of maximum variability in the original data

147 multivariate datasets: principal component 1 is the direction of maximum variance
148 in the multidimensional cloud of points and additional axes are generated to explain
149 the maximum remaining variance while remaining perpendicular to all other axes.

150 In practice, PCA solutions are not derived graphically but are instead calculated
151 using a linear algebra technique involving spectral decomposition of an association
152 matrix to extract the eigenvectors and eigenvalues (these two terms will be
153 explained below). The association matrix defines the interrelationships between the
154 samples based on the covariance or correlation between variables (covariance and
155 correlation are mathematical metrics of the degree to which changes in two vari-
156 ables are associated) (Quinn and Keough 2002). It is important to note that PCA
157 with the covariance matrix will often give significantly different results from
158 PCA with a correlation matrix; however, because PCA is not appropriate for
159 ecological data analysis, the details are not important for this chapter. In brief, the
160 correlation matrix is calculated from standardized data, removing the effects of
161 differences in the variances of the original variables (Quinn and Keough 2002).
162 As such, it is useful for analyzing variables measured in different units or with dif-
163 ferent scales, whereas the covariance matrix (sensitive to differences in variance) is
164 more appropriate when differences in the variances have biological significance.

165 Regardless of the choice of association matrix, PCA will produce a final plot with
166 several principal component axes (“eigenvectors”) each explaining a certain amount
167 of the variance present in the initial data (quantified by the axis “eigenvalue”).
168 Because the principal component axes are linear regressions through the data cloud,
169 each axis can be defined by a linear combination of the analyzed variables. For
170 example,

$$171 \text{PC1} = c_1X_1 + c_2X_2 + \dots + c_nX_n$$

172 The coefficients c_n are termed “loadings” and quantify the importance of each
173 variable in influencing the sample’s position along the principal component axis. In
174 the case of community analysis, they indicate the degree to which a sample’s posi-
175 tion is influenced by the abundance of each species (X_n). When the loading of a
176 species on a particular principal component is high, variations in its abundance are
177 a strong control on the distribution of samples along that axis.

178 Two PCA ordinations of Mistaken Point assemblages, one using a covariance
179 matrix and one using a correlation matrix, are shown in Fig. 1.2 with the eigenval-
180 ues (given the Greek letter λ) displayed by each principal component axis and the
181 loadings of each species displayed as lines radiating from the origin. The graphs are
182 presented solely to demonstrate the interpretation of ordination plots, even though
183 it is likely that neither result accurately reflects the underlying ecological structure
184 of Mistaken Point communities due to the inappropriate linear assumption inherent
185 to PCA. The units on each axis depend on the scale of original measurements and
186 are not of importance for data interpretation. Note that the amount of variance
187 explained by each principal component is low (no more than 40%) – a result that is
188 typical of ecological data ordination. Explained variance decreases with successive
189 principal components so only the first two, or sometimes three, axes are typically

1 Ordination Methods and the Evaluation of Ediacaran Communities

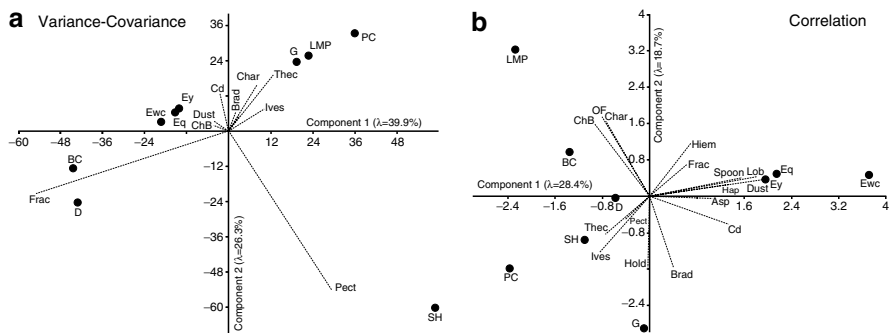


Fig. 1.2 Principal Components Analysis (PCA) ordination of Mistaken Point data (only PC1 and PC2 are shown for simplicity). (a) Ordination using the variance-covariance matrix. Samples are indicated by *black circles* and taxon loadings are indicated by *dotted lines* with the length of the *line* indicating the degree of loading on each axis. (b) Ordination using the correlation matrix

interpreted in ecological studies, although it is possible that lower order axes may also contain important information. In the covariance analysis (Fig. 1.2a), *Fractofusus* (“spindles”) and *Pectinifrons* (“pectinates”) have high loadings (indicated by longer lines radiating from the origin) on axes 1 and 2, primarily because they account for 57% of the variance in the original data. This indicates that variations in their abundance are the primary controls on the position of samples along those axes – for example, principal component 1 is essentially the abundance of *Fractofusus*, from high abundance at negative PC1 scores to low abundance at positive PC1 scores. Variations in the abundance of rare species (*Aspidella*, lobate discs, *Hapsidophyllas* (“networks”), “holdfast fronds”, etc.) make essentially no contribution to the structure of the ordination plot. In contrast, all species contribute equally to the correlation PCA due to the effects of variable standardization (small variations in the length of the loadings are an artifact of displaying only two axes of a multidimensional diagram), which gives as much weight to the abundant *Fractofusus* as it does to *Hiemalora*, only present on three surfaces and never accounting for more than 0.5% of the community.

As mentioned above, it is likely that neither PCA ordination accurately represents the actual ecological structure of Mistaken Point communities. That error arises because PCA requires the assumption of an underlying linear relationship between variables (Euclidean distance) – a severe problem that results in a distorted representation of the original ecological gradient when applied to community data (Gauch and Whittaker 1972; Minchin 1987). Many studies have documented curved unimodal variations in taxon abundance along ecological gradients (depth, altitude, etc.) and a non-linear relationship between compositional similarity and distance between samples along the gradient, with the rate of decrease in similarity reduced at greater distances (Minchin 1987). Because PCA attempts to fit this non-linear relationship to an underlying assumption of linearity, the resulting PCA ordination is distorted and likely does not reflect the actual ecological gradients. One common distortion is the characteristic “horseshoe” shape, in which the ends

219 of the gradient curve inwards are depicted as having greater similarity than is
220 actually the case. This contrasts to other ordination techniques that are not based on
221 the assumption of a linear relationship, such as detrended correspondence analysis
222 (discussed in more detail below), which do not exhibit the horseshoe effect. The
223 effect is less pronounced when beta diversity (between-sample species difference)
224 is low, but when multiple gradients are present in the original sample space the
225 distortion is severe and undetectable, and the results cannot be interpreted (Fasham
226 1977; Minchin 1987). As a result, PCA should not be used for ordination of eco-
227 logical data.

228 **1.4.2 Correspondence and Detrended Correspondence** 229 **Analysis (CA/DCA)**

230 The recognition of faults in PCA methodology when applied to community data
231 (e.g., Gauch and Whittaker 1972) led to the development of new techniques for
232 ecological ordination. Correspondence analysis, the underlying mechanism for DCA,
233 is an eigenanalysis technique similar to principal components analysis (CA can
234 also be calculated through an iterative process called reciprocal averaging) (Hill
235 and Gauch 1980). Whereas PCA conceptually seeks to rotate axes through the
236 multidimensional cloud of sample points to maximize the variance explained, CA
237 can be thought of as rotating the axes simultaneously through both species and
238 sample points to maximize the correspondence between the two. In matrix algebra
239 terminology, this simultaneous axis rotation means that eigenvectors for species
240 and samples are extracted jointly and allows samples and species to be plotted
241 directly on the same axis instead of separately determining loadings of species onto
242 sample axes. Correspondence analysis performs the same matrix decomposition
243 techniques as PCA but on the original data after data transformation to yield
244 chi-square distances rather than on an association matrix of correlation or covari-
245 ance between sites (Chardy et al. 1976; Faith et al. 1987). As a result, many features
246 of the output of CA are analogous to those in PCA, only differing due to the use of
247 chi-square distance rather than Euclidean distance (Kenkel and Orloci 1986;
248 Faith et al. 1987). The sum of CA eigenvalues measures the lack of independence
249 between species and samples in the original data set (the overall chi-squared statistic
250 divided by total frequency – called “inertia”); in PCA they measure the total vari-
251 ance in the original data set. As in PCA, successive axes account for progressively
252 less inertia, with the first axis explaining a relatively high proportion of the lack of
253 independence between species and samples.

254 This chi-square transformation of the original data matrix is a key aspect of CA
255 and is one reason why it is more suitable for ordination of ecological data (Faith
256 et al. 1987). Chi-square distance does not assume a linear relationship between
257 sample dissimilarity and ecological distance, an assumption inherent in PCA that is
258 rarely met by ecological data (Faith et al. 1987; Minchin 1987). As a result, CA
259 correctly ordines samples from the ends of the environmental gradient along the

primary axis, avoiding the “horseshoe effect” common in PCA. The chi-square distance has other limitations, however, and is not robust to variations in the underlying structure of the ecological gradient when taxon abundances deviate from a unimodal normal distribution (Faith et al. 1987), a relatively common feature in natural ecosystems (Oksanen and Minchin 2002). It tends to overweight species with low abundance, exaggerating the distance of samples containing more rare taxa (Minchin 1987), although different software packages contain options for downweighting their importance (Hill and Gauch 1980; Holland et al. 2001; Scarponi and Kowalewski 2004). Chi-square distance also does not reach a constant maximum when samples with no compositional overlap are compared, instead varying depending on the relative occurrence of common and rare species in those samples (Minchin 1987).

Despite the limitations of the chi-square distance, correspondence analysis often performs well at reconstructing the primary ecological gradient (Gauch et al. 1981; Olszewski and West 1997), although there are two potential artifacts of the CA process that may hamper interpretation. First, samples are often compressed at either end of the gradient relative to the middle (Hill and Gauch 1980) because taxon breadths along the gradient are artificially truncated and narrowed at the edge of the study area (Peet et al. 1988). Second, and more significantly, CA does not produce additional axes that are independent of prior axes (although they must not be linearly correlated, they can have a non-linear relationship) and the second axis often appears as a quadratic function of the first (due to projection of a multidimensional dataset into two dimensions) – termed the “arch effect” (Hill and Gauch 1980). Two separate techniques have been devised to correct for these artifacts in CA: detrending to eliminate the arch effect on higher-order axes, and rescaling to reduce compression at the ends of axes. The resulting ordination is called detrended correspondence analysis (DCA) (Hill and Gauch 1980).

Detrending uses a running window to divide the primary axis into segments; all points within a segment are centered by subtracting the mean axis 2 score in that segment from each axis 2 value (Fig. 1.3) (Hill and Gauch 1980). This technique

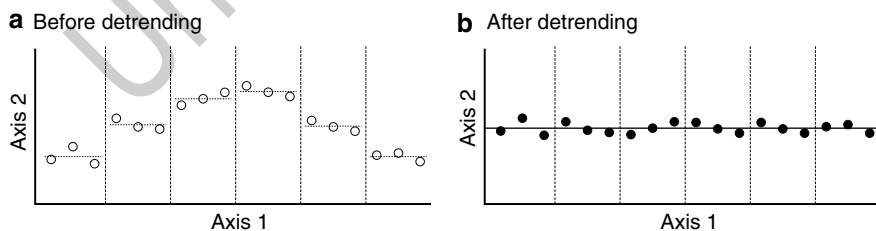


Fig. 1.3 Example illustrating the detrending procedure used in DCA. (a) Axis 1 is divided into segments (dashed lines) and the original points in each section (open circles) are centered by subtracting the mean axis 2 value for that segment (dotted line). (b) The resulting detrended points are shown by filled circles. The actual detrending procedure is more complicated because it uses overlapping running segments and is performed in conjunction with rescaling (Modified from Hill and Gauch (1980))

290 removes any arch effect (by distorting the ordination plot), regardless of whether
291 the arch is an artifact or a real feature of the ecological structure (Kenkel and Orloci
292 1986; Minchin 1987; Wartenberg et al. 1987; Olszewski and West 1997). Detrending
293 can also be sensitive to the number of segments in some cases (Jackson and Somers
294 1991), although the default value (26 segments) often improves the ordination (Peet
295 et al. 1988; Knox 1989).

296 DCA also uses a mathematical rescaling process to remove the effects of gra-
297 dient compression near the ends of axes, assumed to be an artifact (Hill and
298 Gauch 1980). It requires that the abundances of all taxa are normally distributed
299 along the gradient with equal variances (i.e., that species composition changes at
300 the same rate along the gradient) and adjusts the position of species along the
301 axis to maintain a constant within-sample variance of species scores at each point
302 (Hill and Gauch 1980; Peet et al. 1988). This has the benefit of rescaling the axis
303 so that distance can be directly interpreted in terms of compositional change
304 (Peet et al. 1988) but has been criticized because of its potentially unjustified
305 assumption of constant rates of taxon turnover (Wartenberg et al. 1987). Species
306 abundances along a gradient are not always Gaussian (Oksanen and Minchin
307 2002) and different species response curves may have different variances
308 (Holland 2005).

309 The choice of whether or not to detrend the data has been intensely debated
310 (Minchin 1987; Wartenberg et al. 1987; Peet et al. 1988; Knox 1989) and CA
311 (Cisne and Rabe 1978; Olszewski and West 1997) and DCA (Holland et al. 2001;
312 Scarponi and Kowalewski 2004; Holland 2005; Zuschin et al. 2007; Clapham and
313 James 2008) have both been used in paleoecological data analysis. Despite the
314 arbitrary and perhaps unfounded assumptions inherent to DCA (Wartenberg et al.
315 1987), the detrending and rescaling process may yield more accurate and inter-
316 pretable ordination results, especially of long ecological gradients (Peet et al.
317 1988; Knox 1989) or in the case of rapid ecological turnover (Kenkel and Orloci
318 1986), although in other cases they produce distortions in the underlying gradient
319 (Kenkel and Orloci 1986; Minchin 1987). In particular, DCA ordination is often
320 twisted so that residual variation is actually spread over axes 2 and 3, resulting in
321 the characteristic wedge shape, where variation on axis 2 is maximal near the
322 mid-point of axis 1, seen in many DCA plots (e.g., Scarponi and Kowalewski
323 2004; Tomasovych and Siblik 2007; Zuschin et al. 2007; Clapham and James
324 2008). In analyses of simulated data, CA and DCA typically produce accurate
325 ordinations of the primary axis (Gauch et al. 1981) but the structure along axes 2
326 and 3 may be distorted, regardless of the effects of detrending and rescaling. In
327 Mistaken Point data, the detrending/rescaling procedure modifies the spacing
328 between points, increasing the distance between E, G, and BC samples while
329 bringing SH and PC samples closer to the main cluster, but does not greatly alter
330 the relative position of the samples (Fig. 1.4). The main difference is in the posi-
331 tion of the PC sample, which is likely difficult to ordinate due to its low species
332 count (only three taxa). Overall, CA and DCA both appear to result in a reason-
333 able ordination of Mistaken Point surfaces.

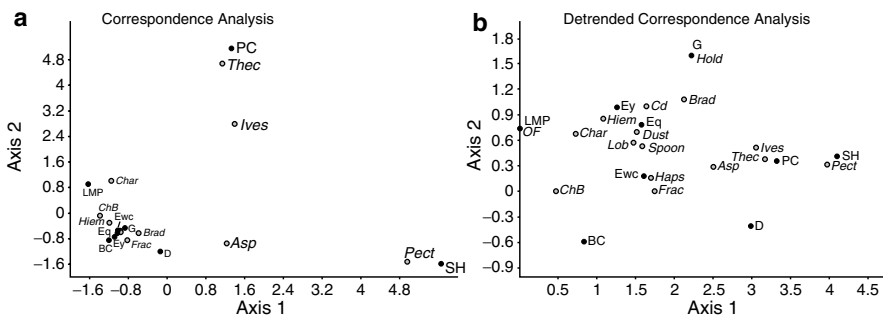


Fig. 1.4 Correspondence Analysis (a) and Detrended Correspondence Analysis (b) ordinations of Mistaken Point data (only the first two axes are shown for simplicity). Sample positions are shown by black filled circles and species positions by gray filled circles

1.4.3 Non-Metric Multidimensional Scaling (NMDS)

334

Eigenanalysis ordination methods such as PCA or DCA are metric techniques that stipulate a proportional relationship between compositional change and distance along the gradient (Kenkel and Orloci 1986; Minchin 1987). As discussed above, ordination results may be distorted when the actual relationship deviates from the idealized assumptions. Another family of techniques compares between-sample similarity using quantitative distance metrics (discussed below) rather than taking an eigenanalysis approach. This section will focus primarily on non-metric multidimensional scaling, the most widespread of these techniques, but it is useful to briefly mention two other distance-based ordination methods: polar ordination and principal coordinates analysis (PCoA).

335
336
337
338
339
340
341
342
343
344

Polar ordination (also called Bray-Curtis ordination) is so named because it projects all samples onto an axis using their relationship to two selected reference points (“poles”), as calculated by the chosen distance metric (Bray and Curtis 1957; Cisne and Rabe 1978; Beals 1984). In the original formulation of Bray and Curtis (1957), the two data points with the maximum separation were chosen as the reference points; later workers proposed a variety of methods for selecting reference points (Beals 1984). Higher-order axes can be constructed from a matrix of residual distances (i.e., the remaining distance not accounted for by the first axis), producing axes that are perpendicular and uncorrelated to the primary axis (Beals 1984). The comparison of all samples to only two reference points, rather than analyzing all intersample differences, may seem like a limiting constraint but polar ordination appears to perform well at ecological ordination (Gauch and Whittaker 1972; Beals 1984).

345
346
347
348
349
350
351
352
353
354
355
356

Principal Coordinates Analysis (PCoA) attempts to maximize the linear correlation between actual intersample distances (using a user-chosen distance metric) and those distances projected onto a two- or three-dimensional space (Gower 1966). It is sometimes called “metric multidimensional scaling” because it is similar to

357
358
359
360

361 non-metric multidimensional scaling but uses a parametric (or “metric”) linear
362 correlation. Although PCoA is able to use an ecologically-appropriate distance
363 metric, the linear correlation imparts similar distortion to that seen in PCA (Podani
364 and Miklós 2002) and PCoA generally performs poorly at ordinating ecological
365 data (Minchin 1987).

366 Non-metric multidimensional scaling differs markedly from those metric approaches
367 by assuming only that the relationship between distance and sample dissimilarity is
368 monotonic (i.e., increasing rank-order distance corresponds to increased rank-order
369 dissimilarity) rather than proportional (Kruskal 1964; Fasham 1977; Kenkel and
370 Orloci 1986; Clarke 1993). In this manner, rank-order NMDS is analogous to non-
371 parametric classical statistics. NMDS uses this rank-order agreement between the
372 calculated solution and the original dataset, a value called “stress,” to assess the quality
373 of ordination results. Analysis begins with a random configuration of points (or a user-
374 specified configuration, such as the results of another ordination method) and performs
375 an iterative optimization procedure to refine the solution until the stress reaches a mini-
376 mum (Clarke 1993). The algorithm may reach a local stress minimum rather than
377 identifying the globally best solution (Fasham 1977; Kenkel and Orloci 1986; Minchin
378 1987; Clarke 1993), but this is rare when the starting configuration is random and can
379 generally be overcome by selecting several random starting points and comparing the
380 final stress to confirm that the optimal solution has been reached (Clarke 1993).
381 The lowest stress indicates the best rank-order agreement between calculated solution
382 and original matrix, with a stress of zero indicating perfect rank-order agreement and
383 stress values less than 0.1 corresponding to a good representation that can reliably be
384 interpreted (Clarke 1993). Higher stress values can still be interpreted, but with greater
385 caution, although stresses greater than 0.2 indicate a poor to nearly random ordering
386 of points (Clarke 1993).

387 Another fundamental difference of NMDS is the ability to select from a variety
388 of similarity or distance coefficients, whereas PCA is based on Euclidean distance
389 and DCA is constrained to use chi-square distance to calculate sample dissimilarity
390 (Faith et al. 1987). This is a significant benefit because Euclidean (and perhaps also
391 chi-square) distance often perform poorly with ecological data (Faith et al. 1987).
392 A myriad of similarity or distance metrics have been proposed, each with benefits
393 and drawbacks for specific data types (Faith et al. 1987; Shi 1993). Studies with
394 various simulated ecological gradients have suggested that the Bray-Curtis,
395 Kulczynski, and Relativized Manhattan (i.e., each species standardized to percent
396 abundance within a sample and normalized to its maximum abundance in any
397 sample) coefficients produce the closest rank-order and linear correlation between
398 compositional dissimilarity and ecological distance along the gradient under most
399 conditions (Faith et al. 1987). Although some coefficients perform better than
400 others with simulated ecological data, there is no objective guide to choosing a
401 distance metric (see Faith et al. (1987) for a detailed review of several distance
402 measures). The Bray-Curtis coefficient was designed for interpretation of ecological
403 data (Bray and Curtis 1957) and performs well on simulated datasets (Faith et al.
404 1987). Because it enjoys widespread use and is intuitive – simply calculate the
405 absolute value of the difference in abundance of a species ($X_i - X_j$) between two

samples divided by the total abundance of that species in the two samples (X_i+X_j), 406
and sum the results for all species – it is a good choice for use in NMDS. It is by 407
no means the only possibility, as other metrics perform well and yield robust 408
solutions, but Bray-Curtis similarity will be used in the NMDS ordination in this 409
chapter. Other distance metrics, such as the Kulczynski coefficient (Faith et al. 410
1987), yield very similar solutions. 411

In contrast to the eigenanalysis methods, NMDS simply arranges the points in 412
 n -dimensional space (where n is a number of dimensions, usually two or three, 413
chosen by the user) rather than extracting multiple orthogonal axes that explain 414
decreasing amounts of variance (Clarke 1993). This has two major implications for 415
viewing and interpreting NMDS ordinations. First, axes in an NMDS ordination 416
have no specific meaning and the data plot can be rotated, translated, or scaled 417
without altering the results, as long as the relative position of the points remains 418
unchanged (Clarke 1993). For example, the ordination is unchanged after rotating 419
points so that the direction of maximum variability is horizontal to facilitate visual 420
comparison with DCA or PCA plots (e.g., Tomasovych and Sibilik 2007). Second, 421
as the NMDS algorithm specifically arranges the samples to fit in n -dimensional space, 422
the best solution for two dimensions will not necessarily be a two-dimensional 423
projection of the best solution for three dimensions (Clarke 1993). This contrasts 424
with eigenanalysis methods, where many eigenvectors are calculated and a plot of 425
axis 1 vs. axis 2 will simply be a two-dimensional projection of the multivariate 426
data cloud. Although it is rare to require more than three dimensions, how should 427
the number of dimensions be chosen? The iteration will yield a better solution 428
(i.e., lower stress) as additional dimensions are included because the original data- 429
set is multivariate, until the number of dimensions is one less than the number of 430
samples. However, a plot of stress against the dimensionality (called a “scree plot,” 431
available in some software packages) will often exhibit a distinct break in slope, 432
providing a rough guideline for the minimum number of dimensions that should be 433
analyzed (Kruskal 1964). It is also reasonable to interpret one extra dimension if 434
the ordination results in additional structure (Kruskal 1964). One caveat: the resulting 435
ordination plot may be distorted when the specified dimensionality is greater than 436
the actual number of dimensions in the original environmental gradient (Austin 1976). 437

NMDS was adopted slowly due to its intensive computational requirements but 438
the calculations are now routine and it is used widely in paleoecological analysis 439
(Olszewski and West 1997; Bonuso et al. 2002; Clapham et al. 2003; Dominici and 440
Kowalke 2007; Tomasovych and Sibilik 2007; Zuschin et al. 2007). Many studies 441
have shown that NMDS produces more accurate ordinations than CA/DCA, for 442
simulated ecological data at least (Fasham 1977; Kenkel and Orloci 1986; Minchin 443
1987), although each technique has individual strengths and weaknesses. In particular, 444
NMDS is less susceptible to distortion when beta diversity is high and when there 445
are multiple underlying gradients with different beta diversities (Fasham 1977). 446
The non-metric technique may be less adept than the eigenanalysis techniques at 447
delineating clusters within the data, but may perform better at optimizing relative 448
distance between samples even when there are clusters (Kenkel and Orloci 1986). 449
As discussed more below, NMDS may be of reduced use when sample size is small; 450

451 in analyses with fewer than 30 samples, use of rank-order may result in significant
452 loss of information (Anderson 1971). Overall, NMDS is a robust ordination technique
453 that performs excellently with ecological data.

454 Figure 1.5 shows NMDS ordination plots for Mistaken Point data using the
455 Bray-Curtis similarity coefficient, based on 2D and 3D dimensionality. As discussed
456 above, the Bray-Curtis coefficient was designed for comparison of ecological sam-
457 ples and produces good results with simulated ecological data (Faith et al. 1987).
458 Three different low-stress two-dimensional solutions (arbitrarily chosen from many)
459 yield virtually the same arrangement of points, differing slightly in the position of
460 the three E surface subsamples (Fig. 1.5a–c). In contrast, the three randomly-selected
461 3D solutions are highly erratic (the substantial difference in arrangement does not
462 result from only viewing two dimensions) (Fig. 1.5d–f). The stress value is lower,
463 but recall that stress is always reduced when extra dimensions are added (Kruskal
464 1964). The unpredictable performance of the 3D solution may indicate that the
465 original environmental gradient was two-dimensional (Austin 1976), but could
466 instead result from the small number of samples and low species richness within
467 each sample. Replicate random analyses did not converge upon a repeated solution
468 of lowest stress, suggesting that there was no single configuration of points that best
469 fit the criteria, likely because there were few samples with few shared taxa.

470 1.5 Comparison and Interpretation of Results

471 These results raise an obvious question – which ordination yields the “correct”
472 ecological representation of Mistaken Point communities? As with all natural data-
473 sets, there is no objective way of testing the accuracy of the ordination results
474 because the positions of samples along the underlying environmental gradients are
475 unknown. Analyses of simulated ecological data suggest that DCA and NMDS
476 should yield the most accurate and robust results (Fasham 1977; Kenkel and Orloci
477 1986; Minchin 1987) and, with the exception of the placement of the PC surface,
478 they produce very similar ordination of the data (Fig. 1.6). Note that the NMDS
479 ordination has been rotated from the plots in Fig. 1.5 to highlight the similarity with
480 the DCA plot, and recall that rotations such as this do not alter the results of the
481 ordination. Two primary gradients can be observed: one from samples with abun-
482 dant *Fractofusus* to samples with no *Fractofusus* and also containing more
483 *Charniodiscus* and *Bradgatia* (vertical axis) and one from samples with abundant
484 *Pectinifrons* to samples with abundant *Charnia* and “*Charnia*” B (horizontal axis).
485 Placement of the PC surface is erratic with different methods, almost certainly
486 because that assemblage only contains three taxa (*Thectardis*, *Ivesheadia*, and
487 *Charnia*). DCA ordination places it close to the SH surface because of their shared
488 abundance of *Ivesheadia*, but its position in the NMDS plot closer to the frond-rich
489 LMP community (with *Charnia*), E surface (with *Thectardis*), and G surface seems
490 intuitively more reasonable.

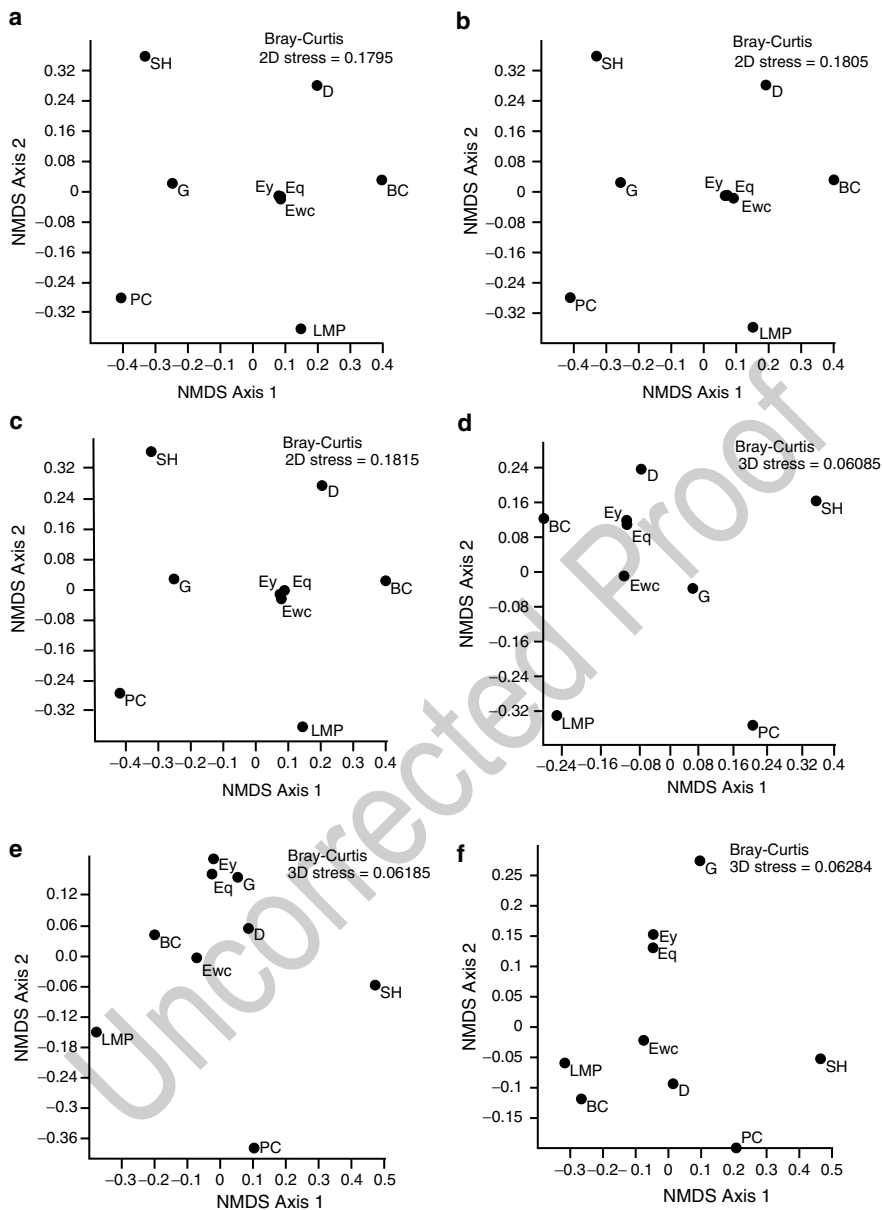


Fig. 1.5 Non-Metric Multidimensional Scaling ordinations of Mistaken Point data. (a–c) Three results of a two-dimensional ordination (arbitrarily chosen) using the Bray-Curtis coefficient. (d–f) Three random results of a three-dimensional ordination using the Bray-Curtis coefficient. Note the instability of the three-dimensional solution compared to the stable two-dimensional results. As discussed in the text, this may indicate that the underlying ecological gradient is two-dimensional or may be a limitation of the small dataset

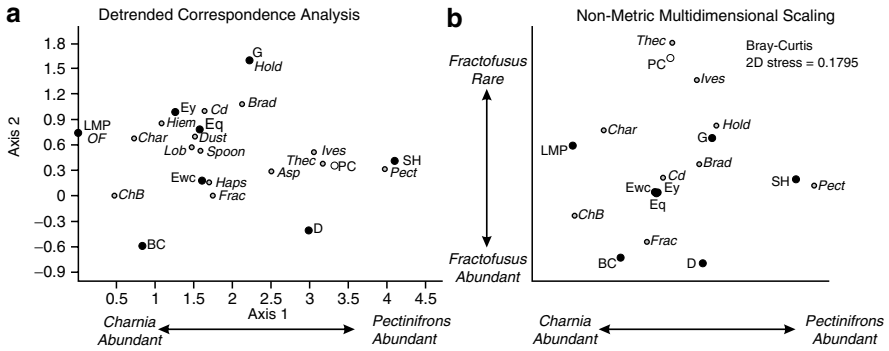


Fig. 1.6 Comparison of DCA and NMDS ordinations of Mistaken Point data, showing interpreted taxonomic gradients responsible for the position of samples. **(a)** Detrended Correspondence Analysis, with samples shown by *black filled circles* (except for the PC surface, which is shown by an *open circle*) and species by *gray filled circles*. **(b)** Non-Metric Multidimensional Scaling (two dimensions, Bray-Curtis coefficient) rotated to align with the DCA results. Samples shown by *black filled circles* except for the PC surface (*open circle*). The positions of selected abundant species are shown by *gray filled circles*; PAST does not ordinate species so their positions were obtained from NMDS ordination in R

491 The next step in an indirect gradient analysis such as DCA or NMDS is inferring
 492 the environmental, evolutionary, ecological, or other gradients that may have been
 493 responsible for generating the taxon gradients identified above. This is most often
 494 accomplished qualitatively by comparing other lines of evidence, for example from
 495 sedimentological interpretation, to the results of the ordination plot, although it is
 496 possible to quantitatively include independent lines of evidence using linear regres-
 497 sion (e.g., Clapham et al. 2003). Although the interpretation is less important than
 498 the methodology for the purposes of this chapter, it may be instructive to briefly
 499 consider possible interpretations. Because Mistaken Point assemblages span >10
 500 Myr of time, there is the possibility that evolutionary changes are an important
 501 control; thus, ordination position may be related to stratigraphic position.
 502 Stratigraphic position is also an indicator of paleoenvironment at Mistaken Point,
 503 shallowing upward from basin plain in the lowest samples (PC, BC) to toe-of-slope
 504 deposits in the Mistaken Point Formation (LMP, D, E, G) to lower slope in the
 505 uppermost sample (SH) (Wood et al. 2003). There is a broad but weak correspon-
 506 dence between axis 1 position (DCA) and stratigraphic position (Fig. 1.6), excluding
 507 the PC surface, likely because the BC and LMP samples predate the first appear-
 508 ance of *Pectinifrons* whereas the SH sample postdates the last appearance of
 509 *Fractofusus*. This implies that evolutionary changes in the Mistaken Point biota
 510 were one of the probable controls on community composition, as expected for
 511 samples spanning such a long time interval. Clapham et al. (2003) argued that each
 512 fossil surface may also reflect a snapshot of a different stage within an ecological
 513 succession trend from *Pectinifrons* and *Fractofusus*-dominated early succes-
 514 sional communities to frond-dominated late successional communities. Mistaken
 515 Point communities have the temporal resolution to preserve a near-instantaneous

snapshot of the community at a point in time; although the snapshots occur 516
 randomly throughout the stratigraphic section. Ordination techniques are ideally 517
 suited to take a jumbled series of snapshots and organize them into a gradient, 518
 whether that gradient is related to ecological succession or some other factor. The 519
 taxonomic gradient inferred for ecological succession can be observed along the 520
 vertical axis 2 in the DCA and NMDS plots, although it is modified by evolutionary 521
 changes in the biota along axis 1 (Fig. 1.6). Other ecological characteristics are at 522
 least consistent with expectations of a succession model (Clapham et al. 2003) but 523
 more surfaces may be needed to strengthen or falsify the link to ecological succession. 524

1.6 Conclusion 525

Quantitative ordination techniques are ideally suited for analyzing multivariate 526
 community ecology data. Principal Components Analysis (PCA) uses Euclidean 527
 distance to relate species dissimilarity to distance along the gradient, requiring the 528
 inappropriate assumption of a linear species response. Because that assumption is 529
 almost always invalid for community data, PCA ordinations are distorted, often 530
 severely, into a horseshoe shape or worse, usually making them uninterpretable. 531
 Although PCA is not suited for ecological ordination, it is still quite appropriate for 532
 data in morphometric or paleobiogeographic analyses, for example, where the 533
 dissimilarity-distance response is closer to linear (Chaps. 2 and 3). Detrended 534
 Correspondence Analysis (DCA) and Non-Metric Multidimensional Scaling 535
 (NMDS) can both yield accurate ecological ordinations. NMDS performs slightly 536
 better on simulated ecological data but DCA may be better at extracting the primary 537
 gradient. Detrending often results in a more accurate ordination of simulated data 538
 than in unaltered Correspondence Analysis (CA), but has also been criticized for its 539
 ad hoc methodology and assumptions. Thus, the best approach for analyzing your 540
 own paleocommunity data depends on the ultimate goal of the study. If you wish to 541
 identify the primary environmental gradient and reconstruct sample and species 542
 placement along that gradient, DCA is an excellent approach. If you are instead 543
 interested in the overall relationship among samples, NMDS may be the best 544
 option. Since DCA and NMDS ordinations have very complementary strengths, the 545
 most robust approach is to apply both methods; the overall result should be similar 546
 and gradients or clusters resolved by both DCA and NMDS are likely to be a good 547
 reflection of the real underlying ecological structure. 548

Acknowledgments Rodrigo Sala and Marc Laflamme assisted with data collection. Fieldwork 549
 in 2000 and 2001 in the Mistaken Point Ecological Reserve was carried out under Scientific 550
 Research Permits granted by the Parks and Natural Areas Division, Department of Tourism, 551
 Culture, and Recreation, Government of Newfoundland and Labrador. The original data collection 552
 was supported by a Natural Sciences and Engineering Research Council of Canada (NSERC) 553
 grant (to Guy M. Narbonne) and by an NSERC postgraduate scholarship (to Clapham). Thoughtful 554
 reviews from John Huntley, Richard Krause, and Amelinda Webb helped improve many aspects 555
 of this contribution. 556

557 **References**

- 558 Anderson AJB (1971) Ordination methods in ecology. *J Ecol* 59:713–726
- 559 Austin MP (1976) Performance of four ordination techniques assuming three different non-linear
560 species response models. *Vegetatio* 33:43–49
- 561 Beals EW (1984) Bray-Curtis ordination: an effective strategy for analysis of multivariate ecological
562 data. *Adv Ecol Res* 14:1–55
- 563 Bonuso N, Newton CR, Brower JC et al (2002) Statistical testing of community patterns: upper-
564 most Hamilton Group, Middle Devonian (New York State: USA). *Palaeogeogr Palaeoclimatol*
565 *Palaeoecol* 185:1–24
- 566 Botquelen A, Gourvennec R, Loi A et al (2006) Replacements of benthic associations in a
567 sequence stratigraphic framework, examples from the Upper Ordovician of Sardinia and
568 Lower Devonian of the Massif Armoricain. *Palaeogeogr Palaeoclimatol Palaeoecol*
569 239:286–310
- 570 Bray JR, Curtis JT (1957) An ordination of the upland forest communities of southern Wisconsin.
571 *Ecol Monogr* 27:325–349
- 572 Chardy P, Glemarec M, Laurec A (1976) Application of inertia methods to benthic marine ecology:
573 practical implications of the basic options. *Estuar Coast Mar Sci* 4:179–205
- 574 Cisne JL, Rabe BD (1978) Coenocorrelation: gradient analysis of fossil communities and its
575 applications in stratigraphy. *Lethaia* 11:341–364
- 576 Clapham ME, James NP (2008) Paleoeecology of Early-Middle Permian marine communities in
577 eastern Australia: response to global climate change in the aftermath of the late Paleozoic ice
578 age. *Palaios* 23:738–750
- 579 Clapham ME, Narbonne GM, Gehling JG (2003) Paleoeecology of the oldest-known animal communities:
580 Ediacaran assemblages at Mistaken Point, Newfoundland. *Paleobiology*
581 29:527–544
- 582 Clarke KR (1993) Non-parametric multivariate analyses of changes in community structure. *Aust*
583 *J Ecol* 18:117–143
- 584 Dominici S, Kowalke T (2007) Depositional dynamics and the record of ecosystem stability: early
585 Eocene faunal gradients in the Pyrenean foreland, Spain. *Palaios* 22:268–284
- 586 Faith DP, Minchin PR, Belbin L (1987) Compositional dissimilarity as a robust measure of ecological
587 distance. *Vegetatio* 69:57–68
- 588 Fasham MJR (1977) A comparison of nonmetric multidimensional scaling, principal components
589 and reciprocal averaging for the ordination of simulated coenoclines and coenoplanes. *Ecology*
590 58:551–561
- 591 Gauch HG, Whittaker RH (1972) Comparison of ordination techniques. *Ecology* 53:868–875
- 592 Gauch HG, Whittaker RH, Singer SB (1981) A comparative study of nonmetric ordinations.
593 *J Ecol* 69:135–152
- 594 Goodall DW (1954) Objective methods for the classification of vegetation. *Aust J Bot*
595 2:304–324
- 596 Gower JC (1966) Some distance properties of latent root and vector methods used in multivariate
597 analysis. *Biometrika* 53:325–338
- 598 Hammer Ø, Harper DAT, Ryan PD (2001) PAST: paleontological statistics software package for
599 education and data analysis. *Palaeontol Electronica* 4:1–9
- 600 Hill MO, Gauch HG (1980) Detrended correspondence analysis: an improved ordination technique.
601 *Vegetatio* 42:47–58
- 602 Holland SM (2005) The signatures of patches and gradients in ecological ordinations. *Palaios*
603 20:573–580
- 604 Holland SM, Miller AI, Meyer DL et al (2001) The detection and importance of subtle biofacies
605 within a single lithofacies: the Upper Ordovician Kope Formation of the Cincinnati, Ohio
606 region. *Palaios* 16:205–217
- 607 Jackson DA, Somers KM (1991) Putting things in order: the ups and downs of detrended correspondence
608 analysis. *Am Nat* 137:704–712

1 Ordination Methods and the Evaluation of Ediacaran Communities

James FC, McCulloch CE (1990) Multivariate analysis in ecology and systematics: panacea or Pandora's box. <i>Annu Rev Ecol Syst</i> 21:129–166	609 610
Jongman RHG, ter Braak CJF, Van Tongeren OFR (1995) Data analysis in community and landscape ecology. Cambridge University Press, Cambridge	611 612
Kenkel NC, Orloci L (1986) Applying metric and nonmetric multidimensional scaling to ecological studies: some new results. <i>Ecology</i> 67:919–928	613 614
Knox RG (1989) Effects of detrending and rescaling on correspondence analysis: solution stability and accuracy. <i>Vegetatio</i> 83:129–136	615 616
Kruskal JB (1964) Multidimensional scaling by optimizing goodness of fit to a nonmetric hypothesis. <i>Psychometrika</i> 29:1–27	617 618
McKinney FK, Hageman SJ (2006) Paleozoic to modern marine ecological shift displayed in the northern Adriatic Sea. <i>Geology</i> 34:881–884	619 620
Minchin PR (1987) An evaluation of the relative robustness of techniques for ecological ordination. <i>Vegetatio</i> 69:89–107	621 622
Oksanen J, Minchin PR (2002) Continuum theory revisited: what shape are species responses along ecological gradients? <i>Ecol Model</i> 157:119–129	623 624
Olszewski TD, West RR (1997) Influence of transportation and time-averaging in fossil assemblages from the Pennsylvanian of Oklahoma. <i>Lethaia</i> 30:315–329	625 626
Palmer MR (1993) Putting things in even better order: the advantages of canonical correspondence analysis. <i>Ecology</i> 74:2215–2230	627 628
Peet RK, Knox RG, Case JS et al (1988) Putting things in order: the advantages of detrended correspondence analysis. <i>Am Nat</i> 131:924–934	629 630
Podani J, Miklós I (2002) Resemblance coefficients and the horseshoe effect in principal coordinates analysis. <i>Ecology</i> 83:3331–3343	631 632
Quinn GP, Keough MJ (2002) Experimental design and data analysis for biologists. Cambridge University Press, Cambridge	633 634
Rodriguez J (2004) Stability in Pleistocene Mediterranean mammalian communities. <i>Palaeogeogr Palaeoclimatol Palaeoecol</i> 207:1–22	635 636
Scarponi D, Kowalewski M (2004) Stratigraphic paleoecology: bathymetric signatures and sequence overprint of mollusk associations from upper Quaternary sequences of the Po Plain, Italy. <i>Geology</i> 32:989–992	637 638 639
Shi GR (1993) Multivariate data analysis in palaeoecology and palaeobiogeography – a review. <i>Palaeogeogr Palaeoclimatol Palaeoecol</i> 105:199–234	640 641
Tomasovych A, Siblik M (2007) Evaluating compositional turnover of brachiopod communities during the end-Triassic mass extinction (Northern Calcareous Alps): removal of dominant groups, recovery and community reassembly. <i>Palaeogeogr Palaeoclimatol Palaeoecol</i> 244:170–200	642 643 644 645
Wartenberg D, Ferson S, Rohlf FJ (1987) Putting things in order: a critique of detrended correspondence analysis. <i>Am Nat</i> 129:434–448	646 647
Wood DA, Dalrymple RW, Narbonne GM et al (2003) Paleoenvironmental analysis of the late Neoproterozoic Mistaken Point and Trepassey formations, southeastern Newfoundland. <i>Can J Earth Sci</i> 40:1375–1391	648 649 650
Zuschin M, Harzhauser M, Mandic O (2007) The stratigraphic and sedimentologic framework of fine-scale faunal replacements in the middle Miocene of the Vienna basin (Austria). <i>Palaios</i> 22:285–295	651 652 653